# Quantitative structure-activity relationship (QSAR) study of biscoumarins derivatives, potential HIV-1 integrase inhibitor

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#### **Abstract**

A series of biscoumarins derivatives against HIV-1 integrase was subjected to quantitative structure-activity relationship (QSAR) analysis. The result of the study showed that the inhibitory activity, as determined in the cell culture model, was highly correlated with the electronic (HOMO) and lipophilic effects exhibited by the substitution (R). The relationship can be expressed by the following regression equation:  $pIC_{50}$ = -1.903( $\pm$ 1.962) + 0.442( $\pm$ 0.066)logP - 4.504( $\pm$ 1.788) $\epsilon_{Lumo}$ .

Key words: HIV-1 integrase, biscoumarins derivatives, QSAR

# 中文摘要

針對具有抑制人類免疫缺失病毒整合酶的雙香豆素衍生物進行化合物結構與活性抑制相關性研究。研究發現:整合酶的活性抑制與填滿電子的分子軌域(HOMO)及親脂性具有高度相關性;並可用多變數迴歸方程式 pIC<sub>50</sub>=-1.903(±1.962)+0.442(±0.066)logP-4.504(±1.788)ε<sub>Lumo</sub>解釋說明此一相關性。

關鍵詞:人類免疫缺失病毒整合酶,雙香豆素衍生物,QSAR

Key words:HIV-1 integrase/ inhibitory activity/ structure-activity relationship/ biscoumarin

## 1. Introduction

Quantitative structure-activity relationship (QSAR) is a tool to rationalize the interaction of chemical compounds with living subject. The idea of QSAR methods is to draw conclusions by likeness assuming that similarity of drugs with respect to certain chemical properties will result in similar biological response. QSAR can be regard as a computer-derived rule which quantitatively describes biological activity in terms of chemical descriptors. Once a QSAR is known, prediction or generation of new compounds with better activity is promising.

Human immunodeficiency virus (HIV) is a lent virus (a member of the retrovirus family) that can lead to acquired immunodeficiency (AIDS). Eventually syndrome most HIV-infected individuals develop AIDS. These individuals mostly die from opportunistic infections or malignancies associated with the progressive failure of the immune system. Anti-HIV (also called antiretroviral) medications are used to control the reproduction of the virus and to slow or halt the progression of HIV-related disease.

Four classes' drugs of anti-HIV medications including non- nucleoside reverse transcriptase inhibitors. nucleoside reverse transcriptase inhibitors, protease inhibitors and fusion inhibitors are approved by the U.S. Food and Drug Administration (FDA) for the treatment of AIDS. When used in combinations, these medications are termed Highly Active Antiretroviral Therapy (HAART), sometimes referred to as a "cocktail". The successful use of HAART can dramatically suppress human immunodeficiency virus 1 (HIV-1) viral replication and effect significant immune reconstitution [1-4]

However, despite full access to antiretroviral agents, the emergence of antiretroviral-resistant HIV-1 strains and/or drug toxicities can spoil effective treatment. Consequently, the need to develop antiretroviral agents with novel mechanisms of action persists for the treatment of HIV-1 infected patients. In October 2007, FDA approved the first drug in the integrase-inhibitor class for the treatment of HIV-1 as part of combination antiretroviral therapy, adding to the available chemotherapeutic agents for the effective treatment of HIV/AIDS.

HIV-1 IN has emerged as an important therapeutic target for the design of anti-HIV agents because the integration is essential for the replication of HIV and the enzyme responsive for integration appears to be absent in the mammalian host. Integrase catalyzes two reactions; 3'-end processing, in which two deoxynucleotides are removed from the 3'-ends of the viral DNA and the strand transfer reaction, in which the processed 3'-ends of the viral DNA are covalently ligated to the host chromosomal DNA [5-7].

Integration of the proviral DNA is a key step in allowing viral DNA to become permanently inserted into the host genome. This step is essential

for the subsequent transcription of the viral genome which leads to production of new viral genomic RNA and viral proteins needed for the production of the next round of infectious virus. Furthermore, integrase uses a single active site to hold two different configurations of DNA substrates, which may restrict the ability of HIV to develop drug resistance to integrase inhibitors. Systematic screening of potential inhibitors has undertaken using mostly purified integrase-based assays. From such screens several integrase inhibitor classes have now been identified [8-12]. Nowadays, natural and synthetic coumarin derivatives represent a wide range of pharmacological prope- rties. They have been reported to be used as antico- agulant [13], antipsoriasis [14], antibiotics [14,15], fun-gicides [16], antioxidant activity [17], anti-inflamm- atory [17], and antitumor agents [18-21]. More recently, coumarin derivatives have

In continuation of our works on the discovery of novel HIV-1 IN inhibitor a quantitative structure-activity relationship (QSAR) study of some active 3,3'-(substituted-benzylidene)-bis-4

been evaluated in the treatment of human

immunodeficiency virus, due to their ability to

inhibit human immunodeficiency virus-1 integrase

(HIV-1 IN) [22,23].

-hydroxycoumarins is reported. In this paper, we tried to analyze the cause of difference of the activity of compounds against HIV-1 IN by using multivariable linear regression analysis and to obtain the information of mechanism of action at molecular level. Physicochemical the and quantum-chemical parameters taken into consideration in QSAR study are log P for hydrophobic effects,  $\varepsilon_{HOMO}$ ,  $\varepsilon_{LUMO}$  as the electronic influences, MR (molar refractivity) for the steric interactions and NRB (number of rotatable bonds) for molecule flexibility.

## 2. Materials and methods

The  $IC_{50}$  (50% inhibitory concentration in  $\mu M$ ) of compounds **1-9** (Table 1) against HIV-1 IN were taken from our previous reported work [22].

The IC<sub>50</sub> data were converted to pIC<sub>50</sub> (negative

logarithmic of  $IC_{50}$  value) for QSAR study. An increase in  $pIC_{50}$  value indicates the enhancement of the activity. The correlations were sought between inhibitory activity and various substituent constants at position R of molecule (**Table 1**).

	ÓH R ÓH		
Comp.	Substituent (R)	${\rm IC}_{50}^{a}$	pIC <sub>50</sub> <sup>b</sup>
1	OCH <sub>3</sub>	3.1	5.509
2	O N OCH3	23.1	4.636
3	CH <sub>3</sub>	96.	4.018
4	CH <sub>3</sub>	23.1	4.636
5	H <sub>3</sub> C CH <sub>3</sub>	102	3.991
6	H <sub>3</sub> C CH <sub>3</sub>	1.8	5.745
7	Br O N O Br	0.96	6.018
8	H <sub>6</sub> C CH <sub>6</sub>	0.58	6.237
9	OH OH	0.49	6.310

<sup>&</sup>lt;sup>a</sup> in vitro IC<sub>50</sub> (50% inhibitory concentration in μM) against HIV-1 IN

The molecular structures of the compounds in selected series were sketched using Chem Draw ultra 10.0 module of CS ChemOffice 2007

molecular modeling software. The sketched structures were then transferred to Chem3D module for generation of three dimensional structure (3D). The geometries of generated 3D structures were

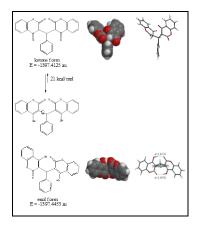
<sup>&</sup>lt;sup>b</sup> negative logarithmic of IC<sub>50</sub> value

pre-optimized using MM2 force field implemented in the Chem3D module of CS ChemOffice2007. These molecular geometries were refined using the quantum chemical program applying package MOPAC 6.0 the PM3 parameterization together with eigenvector following geometry optimi- zation procedure. The gradient norm 0.001 kcal/Å was used to calculate electronic, geometric and energetic parameters for the isolated molecules. The descriptor values for all the molecules were calculated using "compute properties" module of program. Calculated thermodynamic descriptors included critical temperature (T), ideal gas thermal capacity (C), critical pressure (Pc), boiling point (BP), Henry's law constant (H), bend energy (Eb), heat of formation (Hf), total energy (TE), and partition coefficient (PC). Steric descriptors derived were connolly accessible area (CAA), connolly molecular area (CMA), connolly solvent excluded volume (CSEV), exact mass (EM), molecular weight (MW), principal moment of inertia-X component (PMI-X), principal moment of inertia-Y component (PMI-Y), principal moment of inertia-Z component (PMI-Z), number of rotatable bonds (NRB), molar refractivity (MR), and Ovality (OVAL). Electronic descriptors such as dipole (DPL), electronic energy (ElcE), highest occupied molecular orbital energy (ε<sub>HOMO</sub>), lowest unoccupied molecular orbital energy  $(\varepsilon_{LUMO})$ , repulsion energy (RE), VDW-1,4-energy (E14), Non-1,4-VDW energy (E), and total energy (E). Correlation and regression analysis of the QSAR study were run on a PC computer using the Microsoft Excel program. Multiple regression analysis which involves finding the best fit of dependent variable (inhibitory activities against HIV-1 IN) to a linear combination of indepe- ndent variables (descriptors) are used by the least squares method. In the equations, the figures in the parentheses are the standard errors of the regression coefficients, n is the number of compounds, r is the multiple correlation coefficient,  $R^2$  is the determination coefficient,  $Q^2$  is the leave-one-out (LOO) cross vali- dated  $R^2$ , F is the significance test (F-test) and SEE is the standard error of estimate. F-test values are for all equations statistically significant at the 1% level of probability.

## 3. Results and discussion

In organic chemistry, a compound containing a carbonyl group (HC-C=O) is normally in rapid equili- brium with an enol tautomer (C=C-OH). In coumarin molecule the keto-enol tautomerism is possible. Co- mparison the global minimum conformation energy (by Hartree- Fock/6-31G\* calculation) between keto and enol structure of coumarin revealed enol form is more stable than keto form with an energy barrier of 21 kcal/mol (**Figure 1**). The two hydrogen bonds between the hydrogen of enol and the oxygen of lactone in the *bis*-coumarin molecule may contribute the stable conformation.

**Fig. 1** Molecular models of the minimum-energy conformation of *bis*-coumarin



The values of physicochemical parameters and frontier molecular orbital energy levels for compounds are given in **Table 2.** The inhibitory activities of studied compounds against HIV-1 IN are reported with the  $pIC_{50}$  (log 1/C) values. C is the molar concentration of the observed HIV-1 IN inhibitory activity.

Generally, the number of compounds should be greater than four times the number of descriptors in order to get a reliable model using multiple linear regression [23,24]. In this study, we reduced down our pool of descriptors using principal components analysis for building multilinear QSAR models. The treatment started with the reduction in the number of molecular descriptors. If two descriptors intercorrelated highly with each other, then only one of them was selected; descriptors with insignificant variance for the data set treated were also rejected. This helps to speed up the descriptor selection and reduce the probability of including unrelated descriptors by chance. Therefore, log P, lowest unoccupied molecular orbital energy (ELIMO), highest occupied molecular orbital energy ( $\varepsilon_{\text{HOMO}}$ ), number of rotatable bonds (NRB) and molar refractivity (MR) were derived for the multivariable regression analysis

The descriptors derived for mode- ling inhibitory activity of biscoumarins are summarized in **Table 2**. The correlation between the different physic-cochemical descriptors as independent variable and the  $pIC_{50}$  as dependent variable was determined. Some of the statistically significant models are given below and their statistical measures are listed in **Table 3**.

# Model 1

pIC<sub>50</sub>= 2.942(  $\pm$  0.515) + 0.391(  $\pm$  0.084)log P (1)

Model 1 is a monoparametric equation modeled for HIV-1 IN inhibitory activity and has good correlation between biological activity and descriptor as indicated by R=0.87 and explains 76% variance in inhibition. Low standard deviation of the model demonstrates accuracy of the model. The model showed overall significance level better than 99%, with the  $F_{(1,7)}=21.97$  against values of 99% significance  $F_{(1,7,\alpha)}=12.25$ . The p-value is 0.0022 (<0.01), which shows there is significant relationship between log P and biological activity. Leave-one-out (LOO) cross validated value ( $Q^2=0.634$ ) reflects the good predictive power of the model. The plot of experimental vs. predicted property is shown in **Figure 2(a)** 

### Model 2

$$\begin{split} pIC_{50} &= 2.139(\pm 1.060) + 0.279(\pm 0.155)log \ P + 0.008 \\ (\pm 0.009) \ MR \quad &(2) \end{split}$$

Model 2 is a biparametric (log P and MR) equation modeled and was considered to be a poor predictor of activity because of its poor validation (Q<sup>2</sup> = 0.151) although the coefficient of determination (R<sup>2</sup> = 0.786) is higher as compared with model 1. Further, the descriptors log P and MR are found to be highly correlated to each other as shown in the correlation matrix (**table 6**). The correlation coefficient between log P and MR is 0.836 which is outside the acceptable range (< 0.8). However, if the descriptor MR is replaced by NRB, Model 3 is derived.

#### Model 3

 $pIC_{50}$ = 3.673(±0.527) + 0.495(±0.081)log P - 0.135 (±0.061)NRB (3)

Where NRB is Number of Rotatable Bonds. The replacement of MR by NRB drastically improves the quality of prediction as indicated by much better statistical parameters in **Table 3** ( $R^2 = 0.868$ ,  $Q^2 = 0.690$ ). A plot of experimental vs predicted property is shown in **Figure 2(c)**. The negative coefficient of NRB revealed that the molecular flexibility of

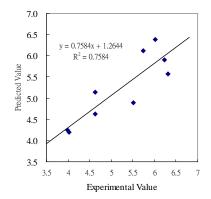
biscoumarins is detrimental to the activity.

#### Model 4

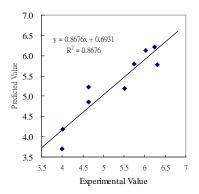
$$\begin{split} pIC_{50} &= \text{-}1.903(\pm 1.962) + 0.442(\pm 0.066)log P - 4.504 \\ &(\pm 1.788) \; \epsilon_{Lumo} \quad \text{(4)} \end{split}$$

Model 4 is the best model since it shows best correlation coefficient R=0.94 and explains 88% variance in inhibition. Further, smaller standard error of estimate, higher F and leave-one-out (LOO) cross validated value ( $Q^2=0.768$ ) demonstrates satisfactory predictive ability of the model. The statistical parameters of Model 4 are shown in **Table 3.** This model indicates that  $\epsilon_{Lumo}$  are highly correlated to the activity. The negative sign of the

**Fig. 2(a)**.Correlation between the predicted  $pIC_{50}$  and the experimental  $pIC_{50}$  by Eq(1)

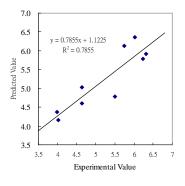


**Fig. 2(c)**.Correlation between the predicted  $pIC_{50}$  and the experimental  $pIC_{50}$  by Eq(3)



coefficient of  $\varepsilon_{Lumo}$  indicates that inhibitory activity increases as  $\varepsilon_{Lumo}$  decreases. The distribution of the HOMO orbitals in the coumarins **8** is shown in the **Figure 3**. By coumarin molecule HOMO orbital is located on the  $\pi$  bond of pyran ring. Atoms with the highest density of this orbital could be the place of the highest reactivity of the substance and the compound could react there as a nucleophilic reagent according to frontier molecular orbital theory. The correlation matrix, experimental and predicted values of activity data are shown in **Table 4** and **5**, respectively, the plot of experimental vs. predicted property is shown in **Figure 2(d)**.

**Fig. 2(b)**.Correlation between the predicted pIC<sub>50</sub> and the experimental pIC<sub>50</sub> by Eq(2)



**Fig. 2(d)**.Correlation between the predicted pIC<sub>50</sub> and the experimental pIC<sub>50</sub> by Eq(4)

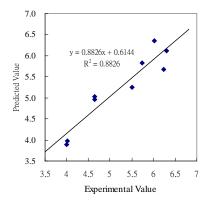


Table 2. QSAR parameters and biological activities.

Compd No.	IC <sub>50</sub> (μM)	pIC <sub>50</sub>	logP	$\epsilon_{Lumo}$	$\epsilon_{Homo}$	NRB <sup>a</sup>	$MR^b$
1	3.1	5.509	4.99	-1.1	-8.91	7	151.162
2	23.1	4.636	4.312	-1.1	-8.52	7	153.111
3	96	4.018	3.219	-0.99	-9.09	8	135.3
4	23.1	4.636	5.604	-0.99	-9.06	9	160.514
5	102	3.991	3.343	-0.96	-8.73	12	158.709
6	1.8	5.745	8.113	-0.92	-8.3	14	209.137
7	0.96	6.018	8.796	-0.97	-8.38	14	214.3
8	0.58	6.237	7.575	-0.94	-9.29	9	186.086
9	0.49	6.310	6.716	-1.12	-9.25	9	232.034

<sup>&</sup>lt;sup>a</sup>NRB=Number of Rotatable Bonds; <sup>b</sup>MR=Molar Refractivity

Fig. 3 LUMO molecular orbital topology distribution for compound 8.

front view	top view	side view

**Table 3**. Statistical parameters of the 4 best scored equations (number of descriptor are not exceed 2)

Eq.	$ND^a$	$R^b$	$R^{2c}$	$Q^{2d}$	$F^{e}$	$\mathtt{p}^f$	SEE g
(1)	1	0.871	0.758	0.634	$F_{(1,7)}=21.97$	0.0022	0.486
(2)	2	0.886	0.786	0.151	$F_{(2,6)}=10.99$	0.0098	0.495
(3)	2	0.931	0.868	0.690	$F_{(2,6)}=19.65$	0.0023	0.388
(4)	2	0.939	0.883	0.768	$F_{(2,6)}=22.56$	0.0016	0.366

<sup>a</sup>ND, <sup>b</sup> R, <sup>c</sup> R<sup>2</sup>, <sup>d</sup> Q<sup>2</sup>, <sup>e</sup> F, <sup>f</sup> p, <sup>g</sup> SD. are the number of descriptors, correlation coefficient, coefficient of determination, leave-one-out(LOO)cross validated R<sup>2</sup>, Fisher-ratio shows the significance of the equations, probability factor and standard error of estimate, respectively.

Table 4. Experimental and predicted pIC<sub>50</sub> values

Compd	nIC (Evn)		pIC <sub>50</sub> (	Pred.)			Resi	due <sup>a</sup>	
No.	$pIC_{50}(Exp)$	Eq(1)	Eq(2)	Eq(3)	Eq(4)	Eq(1)	Eq(2)	Eq(3)	Eq(4)
1	5.509	4.896	4.774	5.197	5.258	-0.613	-0.735	-0.312	-0.251
2	4.636	4.630	4.601	4.861	4.958	-0.006	-0.036	0.225	0.322
3	4.018	4.202	4.150	4.186	3.979	0.185	0.132	0.168	-0.038
4	4.636	5.136	5.022	5.231	5.034	0.500	0.385	0.594	0.397
5	3.991	4.251	4.377	3.707	3.899	0.260	0.385	-0.284	-0.092
6	5.745	6.118	6.121	5.797	5.827	0.374	0.376	0.052	0.082
7	6.018	6.386	6.354	6.135	6.354	0.368	0.336	0.117	0.337
8	6.237	5.908	5.782	6.206	5.679	-0.329	-0.455	-0.031	-0.557
9	6.310	5.572	5.920	5.781	6.111	-0.738	-0.390	-0.529	-0.199

<sup>&</sup>lt;sup>a</sup> Residue= Pred. − Exp.

**Table 5**. Experimental and LOO predicted pIC<sub>50</sub> values

Compd	nIC (Evn)		pIC <sub>50</sub>	(Pred.)				Resi	due <sup>a</sup>	
No.	pIC <sub>50</sub> (Exp)	Eq(1)	Eq(2)	Eq(3)	Eq(4)	_	Eq(1)	Eq(2)	Eq(3)	Eq(4)
1	5.509	4.800	4.532	5.092	5.157		-0.709	-0.977	-0.416	-0.352
2	4.636	4.629	4.549	4.934	5.101		-0.007	-0.087	0.297	0.464
3	4.018	4.289	4.248	4.265	3.957		0.272	0.230	0.247	-0.061
4	4.636	5.198	5.134	5.311	5.090		0.561	0.498	0.675	0.454
5	3.991	4.360	4.663	3.064	3.826		0.368	0.672	-0.927	-0.165
6	5.745	6.249	6.195	5.831	5.874		0.504	0.451	0.086	0.129
7	6.018	6.599	6.456	6.236	6.550		0.581	0.438	0.218	0.532
8	6.237	5.829	5.644	6.187	5.484		-0.407	-0.593	-0.049	-0.753
9	6.310	5.460	4.479	5.658	5.931		-0.850	-1.831	-0.652	-0.379

<sup>&</sup>lt;sup>a</sup> Residue= LOO Pred. – Exp.

## 4. Conclusion

QSAR analysis was performed on a series of biscoumarins using molecular modeling program Chemoffice 2007. QSAR models were proposed for inhibitory activity of the biscoumarins using chemSAR descriptors employing sequential

multiple regression analysis method. The selected models were checked for multicollinearity and correlation with F-test value. The predictive power of each model was validated with determination coefficient R2 method and leave—one—out cross validation Q2 method. It was observed from the best model that biological activity of biscoumarin

derivatives are governed by electronic and lipophilic properties of the molecules. The models also provide valuable insight into the mechanism of action of these compounds. The result of the study suggests involvement of LUMO (biscoumrins)-HOMO (HIV-1 IN) interaction in the mechanism of inhibitory action. The finding of the study will be helpful in the design of potent inhibitory analogs of biscoumarin.

# 5. References

- [1] B. Autran, G. Carcelain, T.S. Li, C. Blanc, D. Mathez, R. Tubiana, et al. Positive effects of combined antiretroviral therapy on CD4+ T cell homeostasis and function in advanced HIV disease. Science. 277, 112-116(1997).
- [2] N.G. Pakker, D.W. Notermans, R.J. de Boer, M.T. Roos, F. de Wolf, A. Hill, et al. Biphasic kinetics of peripheral blood T cells after triple combination therapy in HIV-1 infection: a composite of redistribution and proliferation. Nat Med. 4(2), 208-214(1998).
- [3] J.K. Wong, H.F. Gunthard, D.V. Havlir, Z.Q. Zhang, A.T. Haase, C.C. Ignacio, et al. Reduction of HIV-1 in blood and lymph nodes following potent antiretroviral therapy and the virologic correlates of treatment failure. Proc Natl Acad Sci.; 94(23), 12574-12579(1997).
- [4] Z.Q. Zhang, D.W. Notermans, G. Sedgewick, W. Cavert, S. Wietgrefe, M. Zupancic, et al. Kinetics of CD4+ T cell repopulation of lymphoid tissues after treatment of HIV-1 infection. Proc Natl Acad Sci.;95(3), 1154 -1159(1998).
- [5] R.A. Katz, A.M. Skalka, The retroviral enzymes. Ann. Rev. Biochem., 63, 133-173(1994).
- [6] P. Rice, R. Craigie, D.R. Davies, Retroviral integrases and their cousins, Curr. Opin. Struct. Biol., 6, 76-83(1996).

- [7] C.M. Farnet, B. Wang, J.R. Lipford, F.D. Bushman, Differential inhibition of HIV-1 preintegration complexes and purified integrase protein. Proc. Natl. Acad. Sci. U.S.A., 93, 9742-9747(1996).
- [8] M.R. Fesen, Y. Pommier, F. Leteurtre, S. Hiroguchi, J. Yung., K. W. Kohn, Inhibition of HIV-1 integrase by flavones, caffeic acid phenethyl ester (CAPE) and related compounds. Biochem. Pharmacol, 48, 595-608(1994).
- [9] R.L. LaFemina, P.L. Graham, K. LeGrow, J. C Hastings., A. Wolfe, S.D. Young, E.A. Emini, D.J. Hazuda, Inhibition of human immunodeficiency virus integrase by bis-catechols. Antimicrob. Agents Chemother., 39, 320-324(1995).
- [10] M.R. Fesen, K.W. Kohn, F. Leteurtre, Y. Pommier, Inhibitors of human immunodeficiency virus integrase., Proc. Natl. Acad. Sci. U.S.A., **90**, 2399 -2403(1993).
- [11] R. Dupont, L. Jeanson, J. F. Mouscadet, P. Cotelle, Bioorg. Synthesis and HIV-1 integrase inhibitory activities of catechol and bis-catechol derivatives. Bioorg. Med. Chem. Lett., 11, 3175-3178(2001).
- [12] D. J.Hazuda, P. Felock, M. Witmer, A. Wolfe, K. Stillmock, J.A. Grobler, A. Espeseth, L. Gabryelski, W. Schleif, C. Blau, M.D. Miller, Inhibitors of Strand Transfer That Prevent Integration and Inhibit HIV-1 Replication in Cells. Science, 287, 646-650(2000).
- [13] D. Egan, R. O'Kennedy, E. Moran, D. Cox, E. Prosser, R.D. Thornes, The pharmacology, metabolism, analysis, and applications of coumarin and coumarin-related compounds. Thornes, Drug Metab. Rev. 22, 503-529(1990).
- [14] A. Maxwell, The interaction between coumarin drugs and DNA gyrase. Mol. Microbiol. 9, 681-686 (1993).
- [15] O.A. Abd-Allah, Synthesis and biological

studies of some benzopyrano[2,3-c]pyrazole derivatives. Farmaco 55, 641-649(2000).

[16] A.M. El-Agrody, M.S.A. El-Latif, N.A. El-Hady, A.H. Fakary, A.H. Bedair, Heteroaromatization with 4-Hydroxycoumarin Part II: So-Synthesis of New me Pyrano[2,3-d]pyrimidines, [1,2,4]triazolo[1,5-c]pyrimidines and Pyrimido[1,6-b]-[1,2,4]triazine Derivatives. Molecules 6, 519-527(2001).

[17] A.A. Emmanuel-Giota, K.C. Fylaktakidou, D.J. Hadjipavlou-Litina, K.E. Litinas, D.N. Nicolaides, J. Heterocycl. Synthesis and biological evaluation of several 3-(coumarin-4-yl)-tetrahydroisoxazole and 3-(coumarin-4-yl)-dihydropyrazole derivatives. J Het. Chem. 38, 717-722(2001).

[18] F.H. Dexeus, C.J. Logothetis, A. Sella, K. Fitz, R. Amato, J.M. Reuben, N.J. Dozier, Phase II study of coumarin and cimetidine in patients with metastatic renal cell carcinoma. J. Clin. Oncol. 8, 325-329(1990).

[19. Z.M. Nofal, M.I. El-Zahar, S.S. El-Karim, Molecules 5, Novel Coumarin Derivatives with Expected Biological Activity. 99-113(2000).

[20] H. Madari, D. Panda, L. Wilson, R. Jacobs, A Unique Microtubule Stabilizing Natural Product that Is Synergistic with Taxol. Cancer Res. 63, 1214-1220(2003).

[21] M. Payá; M.L. Ferrándiz; F Miralles; C. Montesinos; A. Ubeda; M.J. Alcaraz, Effects of coumarin derivatives on superoxide anion generation. Arzneimittel-Forschung 43, 655-658 (1993).

[22. C.C. Chiang, J.F. Mouscadet, H.J. Tsai, C.T. Liu, and L.Y. Hsu, Synthesis and HIV-1 Integrase Inhibition of Novel Bis- or Tetra-Coumarin Analogues. Chem. Pharm. Bull. 55, 1740-1743(2007).

[23] A.K. Sen, M.S. Srivastava, Regression Analysis: Theory, Methods and Applications (Springer- Verlag, New York, 1990). [24] S.K. Kachigan, Statistical Analysis: An Interdisciplinary Introduction to Univariate & Multivariate Methods (Radius Press, New York, 1986)